

Pattern of MICRO-RNA-196A2 Polymorphism among type 2 diabetics

Najia Soomro, Sabir Hussain, Ahsan Kazmi, Sarwat Batool, Syed Naqeeb Ali, Syed Liaquat Ali

Department of Pathology, Al-Tibri Medical College, Isra University Karachi, COMSATS University, Al-Nafees Medical College, Isra University, Islamabad, Pakistan

Objective: To identify and determine the pattern of miRNA-196a2 (rs11614913) polymorphism among type 2 diabetics.

Methodology: This case control study was conducted on 311 individuals comprising 161 type 2 diabetics and 150 healthy controls. Genomic DNA was extracted by using non-enzymatic extraction method. Quality of extracted DNA was evaluated by horizontal gel electrophoresis on 1% agarose. The extracted samples were used to run PCR-RFLP for detecting polymorphism in miR-196a2 gene.

Results: Genotypes found were TT (normal

genotype), TC (variant genotype) and CC (variant genotype). The miRNA variant (TC+CC) indicated a significant association with diabetes (69%) as compared to controls (53.3%) ($\chi^2 = 7.98$, $P = 0.004$).

Conclusion: The miRNA variants (TC+CC) indicated a significant association with type 2 diabetes as compared to controls. The percentage of miRNA-196a2 C allele was greater in patients as compared to controls while the frequency of T allele was less in patients compared to the controls.

Keywords: Type 2 diabetes mellitus, miRNA-196a2, DNA.

INTRODUCTION

Diabetes mellitus (DM) is increasing worldwide and type 2 Diabetes mellitus (T2DM) is a serious health burden.¹ The prevalence of T2DM in Pakistan is 11.77%.² It has been indicated that microRNAs play the role of translational repressors and thus control the selected biological processes involved in T2DM.^{3,4} Therefore, dysregulation in miRNAs has considerable impact upon the glucose metabolism of various tissues in T2DM such as the pancreas, adipose tissue, and liver.⁵ Single nucleotide polymorphisms (SNPs) are important genetic alterations in humans.⁶

SNP belonging to miRNA genes/miRNA binding sites have an ability to alter the affinity of binding of miRNAs to their respective messenger RNA thus affecting the targeted gene expression.⁷ Such kinds of SNPs are known part to play a role in susceptibility to a range of diseases including cancer, cardiovascular disease, osteoporosis, and T2DM.⁸ Previous studies have shown common polymorphisms in miR-146a (rs2910164), miR-128a (rs11888095) and miR-27a (rs895819) contributing to neuropathy sensitivity in T2DM.⁹

High-risk SNPs and their alleles (TT, TC and CC) have appeared unbalanced genotype score dissemination among the type 2 DM samples and controls.⁹ Genetic variation in miRNA genes (regulatory regions, primary miRNA, mature miRNA and precursor miRNA) can contribute to pathogenesis of T2DM by altering the expression and form of target gene and miRNA.¹⁰

Hence, focusing on studies of genetic variations in miRNA or miRNA binding sites will help to understand the pathophysiology of T2DM leading to better health management options.^{11,12}

METHODOLOGY

This case-control study was carried out in the department of Pathology, Al Nafees Medical College and Hospital, Islamabad and COMSATS University from Dec, 2017 to Sep, 2018. The sample size of 161 was obtained by purposive sampling technique. Samples were taken from diagnosed cases of T2DM (according to American Diabetes Association criteria). Both males and females were registered in the study with age range 30 to 70 years. Pregnant females and critically ill patients, with diseases having impact on diabetic status (e.g. diabetic ketoacidosis, end-stage kidney disease etc) were excluded from the study. A written and informed consent was taken from all patients.

Data regarding medical illness and demography was collected from each individual. With the help of 5cc sterile syringe venous blood sample was drawn from each individual and transferred to EDTA tubes. To prevent the coagulation of blood gently inverted EDTA tubes to mix EDTA with blood and sample was stored at 4°C. Extraction of genomic DNA was done from the whole blood by using non-enzymatic salting out method. The stored samples were incubated for 15 minutes at room temperature before starting the procedure. DNA was extracted from the blood samples

by stepwise procedure of non-enzymatic extraction method. 1% agarose gel was used for horizontal gel electrophoresis of Genomic DNA. Electrophoresis was done at 500mA and 95 volts for 30minutes in running buffer (1X TAE)¹³. For visualization of product gel was placed under UV light in gel documentation system and gel images were saved (Alpha-Imager Mini Bucher Biotech, Switzerland).

PCR was performed to detect the polymorphisms in rs11614913 gene. Electrophoresis was done at 460mA and 95 volts for 35minutes in running buffer (1XTAE). Gel products were visualized by using gel documentation system under UV light and to confirm the accurate amplification of DNA products compared with DNA ladder. Gel images were saved for record. RFLP technique was performed for SNP genotyping by using Msp1 restriction enzyme (Thermo Fissure-Scientific, USA) (Fig. 1). For selection of enzymes software NEB cutter V2.0 was used. Amplified product was incubated for 24 hours at 37°C. 5% agarose gel was performed for the separation of digested products and visualized by using UV-transiluminator in gel documentation system. The 100bp DNA ladder was used as quality control to confirm the size of digested fragments.¹⁴

Statistical Analysis: The data were analyzed using SPSS version 20. Pearson’s Chi-square test and Fisher’s exact test were applied for statistical evaluation. $p \leq 0.05$ was taken as significant.

RESULTS

Genotypes found were TT (normal genotype), TC (variant genotype) and CC (variant genotype). The miRNA variant (TC + CC) indicated a significant association with diabetes (69%) as compared to controls (53.3%) ($\chi^2 = 7.98, P = 0.004$). The percentage of miRNA-196a2 C allele was 40.3% in patients and 31.6% in controls.

The percentage of T allele was 59.6% in patients compared to 68.3% control. Patients and control percentage showed a significant difference between T and C allele (OR = 1.4, 95%CI = 1.050 – 2.031, P = 0.02) as shown in the Table 1.

DISCUSSION

This is a well-established fact that T2DM has strong genetic susceptibility.¹⁵ After extensive literature review from various search engines, no prior research was

Table 1: Frequency table of miRNA-196a2 T/C polymorphism in study population.

Genotype	Patients (n = 161)	Control (n = 150)	X ² value	P value
TT	50 (31%)	70 (46.6%)	X ² = 8.06	0.017a
TC	92 (57%)	65 (43.3%)		
CC	19 (125)	15 (10%)		
TT	50 (31% ⁹)	70 (46.6%)	X ² = 7.98	0.004a
TC + CC	111 (69%)	80 (53.3%)		
T allele	192 (59%)	205 (68.3%)		
C allele	130 (40.3%)	95 (31.6%)	P = 0.02a OR = 1.4, 95% CI = 1.050-2.031	

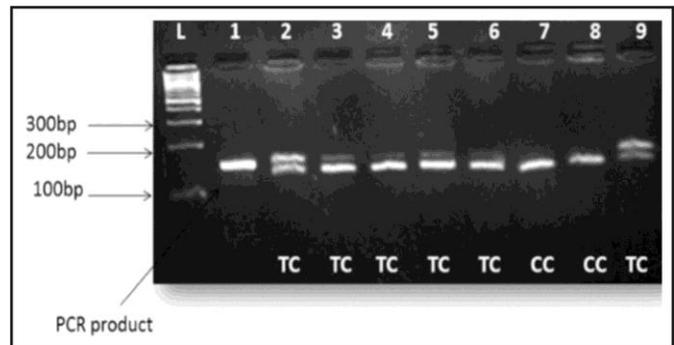


Fig. 1: Identification of the miRNA-196a2 polymorphism. L is 100bp (100 base pair) DNA ladder. Lane 1 is showing PCR product while 2 – 9 are restriction products. Lane 2 – 6 showing TC variant (heterozygous) while lane 7 & 8 showing CC homozygous allele.

found on this topic. However, many studies have highlighted the association of same (microRNA-196a2) polymorphism with various other diseases including cardiovascular diseases, stroke and cancer.¹⁶⁻¹⁸ The present investigation is first of its kind in Pakistan and it aims to find the association of miRNA-196a2 polymorphism at rs11614913 T/C with T2DM as the genetic information of diabetic patients is scarce in Pakistan.

Polymorphism in miRNA-196a2 designated as rs11614913, was the gene observed in the current study. It belongs to the expression of miRNA-196a2. Research shows a long-term workup on this gene to find out its association with various diseases. Ghanbari et al showed that rs11614913: T/C genotype in miR-196a2 was related with waist to hip ratio two target genes were identified.¹⁹ They were experimentally presented as direct targets of miR-196a2. The results suggested that

miRNA-196a2 regulates fat distribution by miRNA-dependent pathway.¹⁹ In another study, Hu et al found that rs11614913: T/C variant genotypes were linked with considerably increased breast cancer risks.²⁰

Titan et al suggested that variant homozygote CC of miR-196a2 rs11614913 was linked with about 25% increased susceptibility to lung cancer as compared to wild-type heterozygote TC and homozygote TT and concluded that SNP rs11614913 in miR-196a2 might increase susceptibility of lung cancer.²¹ XU et al showed that miR-196a2 rs11614913 plays a part in sporadic congenital heart disease susceptibility.²² In a study by Kim et al they tried to find out relationship of miRNA-196a2 with silent brain infarction and ischemic stroke but the connection between silent brain infarction susceptibility and microRNA polymorphisms could not be established.²³

There is one study conducted by Buraczynska et al which concluded that polymorphism in miRNA-196a2 (rs11614913) T/C is associated with an increased risk of cardiovascular disease in patients with type 2 DM. The percentage of T allele was lower in patients with type 2 DM as compared to healthy controls. Frequency of T allele and TT Genotype was high in patients with cardiovascular diseases when compared to those with healthy controls. While frequency of heterozygote TC was same in CVD patients and controls.¹³

However, current study revealed strong evidence on linking role of miRNA-196a2 with type 2 diabetes mellitus. It was found that variant genotypes (TC+CC) showed a significant association with patients of type 2 DM as compared to control group. The percentage of C allele was high in type 2 diabetics while the frequency of T allele was increased in control. This finding may serve in future as basis for screening of type 2 diabetes mellitus in genetically susceptible prediabetes individuals.

CONCLUSION

Our study revealed TT, TC and CC genotypes of microRNA-196a2 are present in type 2 diabetics and healthy individuals. The TC and CC genotypes in microRNA-196a2 (rs11614913) SNP have significant association with type 2 diabetes and hence possible role in screening of susceptible prediabetes. While TT genotype is common among healthy individuals.

Author Contributions:

Conception and design: Sarwat Batool.

Collection and assembly of data: Najia Soomro.

Analysis and interpretation of data: Sabir Hussain.

Drafting of the article: Ahsan Kazmi.

Critical revision of article for important intellectual content: Syed Naqeeb Ali.

Statistical expertise: Syed Liaquat Ali.

Final approval and guarantor of the article: Najia Soomro.

Corresponding author email: Naqeeb: syednaqeeb14@gmail.com

Conflict of Interest: None declared.

Rec. Date: Nov 6, 2020 Revision Rec. Date: Jan 14, 2022 Accept Date: July 22, 2022.

REFERENCES

1. Manonmani M, Manimekalai K. A study of serum magnesium level in Type 2 Diabetes mellitus patients. *J Diab Mellitus* 2018;8:20-26.
2. Meo A, Zia I, Bukhari A and Arain A. Type 2 diabetes mellitus in Pakistan: Current prevalence and future forecast. *J Pak Med Assoc* 2016;66:1637-42.
3. Goda N, Murase H, Kasezawa N, Goda T, Yamakawa-Kobayashi K. Polymorphism in microRNA-binding site in HNF1B influences the susceptibility of type 2 diabetes mellitus: a population based case-control study. *BMC Med Genetics* 2015;16:1-8.
4. You L, Wang N, Yin D, Wang L, Jin F and Zhu Y, et al. Downregulation of long noncoding RNA Meg3 affects insulin synthesis and secretion in mouse pancreatic beta cells. *J Cell Physiol* 2016;231:852-62.
5. Guay C, Roggli E, Nesca V, Jacovetti C and Regazzi R. Diabetes mellitus, a microRNA-related disease? *Transl Res* 2011;157:253-64.
6. López B, Torrent-Fontbona F, Viñas R, Fernández-Real JM. Single Nucleotide Polymorphism relevance learning with Random Forests for Type 2 diabetes risk prediction. *Artif Intell Med* 2018;85:43-9.
7. Cammaerts S, Strazisar M, De Rijk P and Del Favero J. Genetic variants in microRNA genes: impact on microRNA expression, function, and disease. *Front Genet* 2015;6:86-97.
8. Wang X, Li W, Ma L, Gao J, Liu J and Ping F. Association study of the miRNA-binding site polymorphisms of CDKN2A/B genes with gestational diabetes mellitus susceptibility. *Acta Diabetol* 2015;52:951-8.
9. Wu Y, Jing R, Dong Y, Kuang Q, Li Y and Huang Z. Functional annotation of sixty-five type-2 diabetes risk SNPs and its application in risk prediction. *Sci Rep* 2017;7:437-49.
10. Wen Z, Zou X, Xie X, Zheng S, Chen X, Zhu K, Dong S, Liang J, Huang X, Liu D, Wang Y. Association of polymorphisms in miRNA processing genes with type 2 diabetes mellitus and its vascular complications in a southern Chinese population. *Front Endo* 2019;10:461-472.
11. Bhatia P, Raina S, Chugh J, Sharma S. miRNAs: early prognostic biomarkers for Type 2 diabetes mellitus? *Biomark Med* 2015;9:1025-40.

12. Connell T, A Markunas C. DNA methylation and microRNA-based biomarkers for risk of type 2 diabetes. *Curr Diabetes Rev* 2016;12:20-9.
13. Mohammadpour A. Evaluation of a modified salt-out method for DNA extraction from whole blood lymphocytes: A simple and economical method for gene polymorphism. *Pharma Biomed Res* 2018;4:28-32.
14. Gilad S, Lithwick G, Barshack I, Benjamin S, Krivitsky I, and Edmonston B, et al. Classification of the four main types of lung cancer using a microRNA- based diagnostic assay. *J Mol Diagn* 2012;14:510-17.
15. Osman W, Tay GK, Alsafar H. Multiple genetic variations confer risks for obesity and type 2 diabetes mellitus in arab descendants from UAE. *Inter J Obes* 2018;42:1345-53.
16. Buraczynska M, Zukowski P, Wacinski P, Ksiazek K and Zaluska W. Polymorphism in microRNA-196a2 contributes to the risk of cardiovascular disease in type 2 diabetes patients. *J Diab Complications* 2014;28:617-620.
17. Jeon J, Kim J, Kim Y, Oh H, Oh D, Kim J, et al. Association of the miR- 146a, miR-149, miR-196a2, and miR-499 polymorphisms with ischemic stroke and silent brain infarction risk. *Arterioscler Thromb Vasc Biol* 2013;33:420-30.
18. Meiri E, Mueller C, Rosenwald S, Zepeniuk M, Klinke E, Edmonston B, et al. A second-generation microRNA-based assay for diagnosing tumor tissue origin. *Oncologist* 2012;17:801-12.
19. Ghanbari M, Sedaghat S, Looper W, Hofman A, Erkeland J, Franco H, et al. The association of common polymorphisms in mi R-196a2 with waist to hip ratio and mi R-1908 with serum lipid and glucose. *Obes (Silver Spring)* 2015;23:495-503.
20. Hu Z, Liang J, Wang Z, Tian T, Zhou X, Chen J, et al. Common genetic variants in pre- microRNAs were associated with increased risk of breast cancer in Chinese women. *Hum Mutat* 2009;30:79-84.
21. Tian T, Shu Y, Chen J, Hu Z, Xu L, Jin G, et al. A functional genetic variant in microRNA-196a2 is associated with increased susceptibility of lung cancer in Chinese. *Cancer Epidemiol Biomarkers Prev* 2009;18:1183-7.
22. Xu J, Hu Z, Xu Z, Gu H, Yi L, Cao H, et al. Functional variant in microRNA-196a2 contributes to the susceptibility of congenital heart disease in a Chinese population. *Hum Mutat* 2009;30:1231-6.
23. Kim JO, Bae J, Kim J, Oh SH, An HJ, Han IB, et al. Association of microRNA biogenesis genes polymorphisms with ischemic stroke susceptibility and post-stroke mortality. *J Stroke* 2018;20:110-134.